

Forum Review

The Biphasic Nature of Nitric Oxide Responses in Tumor Biology

LISA A. RIDNOUR,¹ DOUGLAS D. THOMAS,¹ SONIA DONZELLI,¹ MICHAEL G. ESPEY,¹
DAVID D. ROBERTS,² DAVID A. WINK¹, and JEFFREY S. ISENBERG²

ABSTRACT

The dual or biphasic responses of cancer to nitric oxide (NO) arise from its concentration dependent ability to regulate tumor growth, migration, invasion, survival, angiogenesis, and metastasis. The outcome of these various NO-dependent processes is dictated by several factors including NO flux, the chemical redox environment, and the duration of NO exposure. Further, it was recently discovered that an NO-induced redox flux in vascular endothelial cells hypersensitizes these cells to the antiangiogenic effects of thrombospondin-1. This suggests a novel treatment paradigm for targeting tumor-driven angiogenesis that combines redox modulation with mimetic derivatives of thrombospondin-1. This article discusses the biphasic nature of NO in cancer biology and the implications of NO-driven redox flux for modulation of tumor-stimulated angiogenesis, growth, and metastasis. *Antioxid. Redox Signal.* 8, 1329–1337.

INTRODUCTION

THE FIELD OF NITRIC OXIDE STUDY has emerged as one of the more prolific areas in biomedical science, as it affects most every aspect of physiology and the medical sciences. Unlike those of bioeffectors whose actions are often based on specific structure–function relations, the biologic properties of NO are ultimately determined by redox chemistry and the location of NO as well as derived reactive nitrogen oxide species (RNOS), which are key determinants of the cellular response to NO. Over the course of the last two decades, studies have demonstrated how these basic chemical reactions regulate normal physiology and pathology (54, 72–74). Although studies have provided new insight into many conditions such as ischemia–reperfusion injury, heart disease, and immunology, this review focuses on the role of NO in cancer biology.

THE DICHOTOMOUS BEHAVIOR OF NO IN CANCER

Nitric oxide (NO) is released intracellularly during the oxidation of L-arginine by the nitric oxide synthase (NOS) enzymes. Three isoforms of NOS have been identified; neuronal (nNOS, NOS1) and endothelial (eNOS, NOS3) are constitutive Ca²⁺-dependent forms of the enzyme that are regulated by negative feedback and release low fluxes of NO over a short period to regulate neural and vascular function, respectively (60). In addition, the Ca²⁺-independent inducible (iNOS, NOS2) form produces large amounts of NO throughout the life of the enzyme and is involved in immune surveillance. The distinct timing and control of these three isoforms dictate the high–low range of NO flux required for specific physiologic processes as well as the generation of other RNOS that facilitate numerous redox-regulated pathways.

¹Tumor Biology Section, Radiation Biology Branch, National Cancer Institute, NIH, Bethesda, Maryland.

²Biochemical Pathology Section, Laboratory of Pathology National Cancer Institute, NIH, Bethesda, Maryland.

The role of NO in cancer has been quite perplexing, as both protective and cytotoxic responses have been demonstrated (24, 42). NO was first shown as an essential component of the macrophage antitumor activity against leukemia cells (24). Recently, tumoricidal activity of NO was further examined by overexpression of iNOS in various tumor cell lines (41). In this study, the transfection of human tumor cells with an adenoviral vector containing iNOS resulted in enhanced iNOS activity 100-fold beyond control. The complete inhibition of tumor growth as well as regional lymph node metastases in nude mice occurred in an iNOS/NO-dependent manner as tumor cells transfected with mutated forms of iNOS cDNA demonstrated reduced tumor-suppressive activity, whereas wild-type (wt) iNOS transfectants yielded complete growth inhibition *in vivo*. Inducible NOS overexpression was also associated with dramatic upregulation of vascular endothelial growth factor (VEGF) and interleukin (IL)-8 angiogenic cytokines, which were associated with enhanced *in vitro* endothelial cell tube formation. Moreover, the ablated tumor growth was associated with NO-mediated cytostasis and enhanced apoptosis (34%) of the transfected cells as well as bystander cells (41, 75–77). Similar NO-induced tumor-suppressive effects were observed in nude mice implanted with malignant rat cells previously exposed to activated Kupffer cells or cell-culture supernatants of activated Kupffer cells (6). In this study, the tumoricidal effects involved upregulation of tumor-necrosis factor (TNF)- α and interferon (IFN)- γ , with concomitant increases in NO production; NO-induced tumor apoptosis was associated with increased DNA damage, elevated caspase-8, and Bcl2 suppression (6).

These effects of acute, high concentrations of NO on tumor growth and metastasis are provocative and warrant a comparison of iNOS and NO levels in normal tissue and human cancer as they relate to conditions of chronic inflammation (20, 26, 73). Toward this end, clinical and experimental studies have demonstrated inverse correlations between iNOS expression and patient survival (12, 38, 65). Another report has demonstrated increased iNOS levels in tumor tissue of colorectal cancer patients (CRCs) when compared with the surrounding normal tissue; beyond this elevated level, iNOS expression declined as a function of tumor progression toward a more metastatic state (1, 20). This observation suggests the occurrence of an NO biphasic response as a function of the levels of iNOS expression and NO concentration over the course of tumor promotion and progression (20). Interestingly, the involvement of an oxidant biphasic response in tumor progression has also been demonstrated (52).

The idea of an NO biphasic response in tumor biology suggests that low NO levels are required for tumor promotion and growth. As the levels of NO increase beyond an "optimal" concentration for tumor growth and survival, growth arrest and/or cell-death pathways are initiated. Cell death may be circumvented by modulation or mutation of oncogenes and/or tumor suppressors (*i.e.*, P53 and Bcl2), resulting in the selective survival and clonal expansion of cell populations that are NO resistant (12, 20, 27). The circumvention of cell death in response to intolerable levels of NO may be supported by the *in vivo* growth results of two human tumor cell lines (253J BV bladder and SKOV3 ipl ovarian cancer cell lines) transfected with adenoviral vector containing iNOS

(41). In these transfectants, the *in vivo* growth of cells expressing high levels of iNOS was suppressed for ~30 days before the observation of small but measurable tumors. Although it is possible that the latent tumor development was associated with loss of the iNOS expression vector, it is also plausible that the resultant tumors represented a population of NO-resistant cells. The elucidation of growth and metastatic potential of these or similarly derived cells would be invaluable to this debate. Toward this end, enhanced *in vivo* growth and metastatic potential has been demonstrated by DLD-1 human colon adenocarcinoma cells that were stably transfected with iNOS (32).

Endothelial NOS is also relevant in tumor progression, as it has been shown to mediate tumor growth and metastasis by stimulation of tumor cell invasion, migration, and angiogenesis in clonal derivatives of spontaneous mammary tumors (38). Further studies demonstrated that NO-induced migration of these tumor cells was cGMP and pERK dependent, suggesting that low NO levels mediated the response (31). Mural cell recruitment is a process required for vascular morphogenesis and maturation. An elegant study by Kashiwagi *et al.* (34) demonstrated that NO produced primarily by eNOS in vascular endothelial cells mediates mural cell recruitment, which occurred during tumor angiogenesis of highly metastatic B16F10 murine melanomas. Moreover, the implantation of B16F10 cells in eNOS^{-/-} and iNOS^{-/-} animals showed a requirement of eNOS in NO-mediated tumor vessel branching and longitudinal extension, but not circumferential growth of blood vessels in B16 melanomas. The results of these studies have demonstrated NO concentration dependencies suggestive of a biphasic role of NO in tumor biology; whereas low levels of NO promote tumor growth, high levels are cytostatic, cytotoxic, and can generate NO-resistant cell populations as well (53).

NO CONCENTRATION DEPENDENCE AND SIGNAL TRANSDUCTION

A key affect of NO is its ability to modulate protein and enzyme activities by direct interaction of NO with Fe²⁺ centers or through protein modification by other RNOS (54). The transcription factors P53 and HIF-1 are important signaling molecules in cancer biology that are modulated by NO and regulate cellular growth, apoptosis, and adaptation to stress through altered gene expression (80). The Brune laboratory has performed extensive studies identifying both concentration- and time-dependent effects associated with NO-induced apoptotic death of macrophages in response to NO donors of differing half-lives (5). These studies have demonstrated NO-mediated apoptosis through the upregulation of P53, activation of caspase enzymes, and modulation of the Bcl2 family of proteins, which occurs independent of cyclic guanosine monophosphate (cGMP) (5). Moreover, NO regulation of these pathways is exquisitely sensitive and dependent on the environmental milieu, as the introduction of other reactive oxygen species (ROS) such as superoxide (O₂⁻) can redirect signaling, resulting in a shift from cell-death to cell-survival pathways, which is also cell specific and glutathione depen-

dent (3, 5, 62, 71). Mechanisms associated with redox suppression of apoptosis involve S-nitrosation of active-site cysteine residues, resulting in the inactivation of key apoptotic enzymes such as caspases (4, 37, 43, 48, 78, 79).

While P53 regulates cell cycle, apoptosis, and DNA repair in relation to stress, HIF-1 responds to subtle changes in oxygen tension resulting from mild hypoxia and is therefore associated with mediation of cell-survival pathways. Interestingly, the intracellular levels of these transcription factors are similarly regulated by posttranslational modifications culminating in proteasomal degradation (56, 81), which are influenced by NO. In an unstressed cell, HIF-1 α is targeted for proteasomal degradation by hydroxylation at Pro 402 and 564 via prolyl hydroxylases and subsequent conjugation with von Hippel-Lindau/E-3 ubiquitin ligase complex. Both NO and mild hypoxia inhibit prolyl hydroxylase function and proteasomal degradation, resulting in HIF-1 α stabilization and accumulation (47, 56, 81). Prolyl hydroxylases are nonheme Fe²⁺-containing proteins; the inhibitory affects of NO are proposed to involve competitive inhibition at the Fe²⁺-active site of the enzyme (47). In addition, cGMP-independent NO stabilization of HIF-1 α and subsequent DNA binding activity by the NO donor DETA/NO (100 and 500 μ M) was mediated by S-nitrosation involving NO⁺ equivalents (49), suggesting the involvement of both NO and RNS in HIF-1 regulation.

Under normoxic conditions, P53 is also maintained at low or undetectable levels via MDM2 conjugation and cytoplasmic proteasomal degradation. Nitric oxide can interfere with the degradation process by causing posttranslational modifications, which stabilize P53 through phosphorylation and acetylation at several sites of the protein (26, 35, 46, 64, 82). Furthermore, cell fractionation and heterokaryon analysis have demonstrated nuclear accumulation of phosphorylated P53 protein after stabilization by NO, suggesting that impaired nuclear export contributes to P53 stabilization in response to NO (57). Therefore, these studies have demonstrated the involvement of several mechanisms in association with NO-mediated stabilization of P53 and HIF-1 α , which are key regulators of cancer progression.

Other signaling cascades important to cell death and survival are the mitogen-activated protein kinases (MAPK), which include extracellular regulated kinase (ERK_{1/2}), c-Jun NH₂-terminal kinase (JNK) and p38; in addition, Akt is a downstream target of phosphatidylinositol 3'-kinase that is important in the transmission of cell-survival signals of growth factors. Chaudhuri and co-workers (50) demonstrated that high concentrations of NO (1 mM DETA/NO) lead to enhanced apoptosis by inactivation of the prosurvival ERK_{1/2} and Akt signaling cascades through enhanced levels of MKP-1 phosphatase in breast cancer cells.

Signal-transduction mechanisms associated with NO concentration, duration of exposure, and NO response as it relates to chronic inflammation and the development of colon cancer also have been reported (26). Nitric oxide from various donors (>100 μ M Sper/NO, DETA/NO, or GSNO) induced both ataxia-telangiectasia mutated (ATM) and ATM-Rad3-related (ATR) kinase-dependent p-ser-15 P53 posttranslational modifications, leading to an increase in p21^{waf-1} and a G₂M cell-cycle arrest (26). Similar levels of p-ser-15 P53 were identified in MCF7 breast carcinoma cells

co-cultured with stimulated ANA-1 murine macrophage expressing iNOS. Therefore, the fluxes of NO produced by the donor at specific concentrations were representative of the microenvironment surrounding iNOS-expressing leukocytes. Moreover, noncancerous colon tissues from patients with ulcerative colitis (a cancer-prone chronic inflammatory disease) demonstrated a positive correlation between the levels of iNOS protein and p-ser-15 P53, as immunostaining of HDM-2 and p21^{waf-1} was consistent with transcriptionally active P53 in these tissues. This study highlights not only a pivotal role of NO in the induction of cellular stress and the activation of a P53 response pathway during chronic inflammation, but also the exploitation of the same pathway by cancer cells as a protective mechanism against inflammatory stress (26).

The effect of NO concentration and duration on posttranslational regulation of signaling proteins under normoxic conditions has also been examined. This work demonstrated discrete NO-threshold profiles associated with pERK, p-ser15-P53, and HIF-1 α expression in MCF-7 breast cancer cells (64). A 30-min exposure to low steady-state concentrations of NO (<50 nM), which corresponded to 1–10 μ M Sper/NO, induced transient ERK_{1/2} phosphorylation that was cGMP dependent. Intermediate Sper/NO concentrations (50 μ M) yielding >100 nM steady-state levels of NO caused HIF-1 α stabilization, which required the persistence of intermediate fluxes of NO. High steady-state fluxes of NO (>300 nM), corresponding to 100 μ M Sper/NO concentration, led to the accumulation of p-ser15-P53. Where HIF-1 α responded to acute increases and decreases of NO, P53 phosphorylation occurred after 2 h of exposure to the NO donor and persisted well after the NO flux disappeared. In contrast, ERK phosphorylation was transient and decreased despite the continued presence of NO. These observations are indicative of the unique temporal and concentration dependence of signaling profiles in response to NO and provide insight into the dichotomous behavior of NO. For example, HIF-1 and pERK are progrowth, whereas P53 phosphorylation is associated with cell-cycle delay, DNA repair, and apoptosis. The ramifications *in vivo* suggest that, despite the toxic effects of high fluxes of NO, these discrete signaling profiles could facilitate the selection and clonal expansion of a mutated genotype(s) under chronic inflammation and iNOS expression (20, 26, 64).

Another important variable of these distinct concentration gradients relative to their biologic response is the enzymatic activity of specific NOS isoforms. Two of the isoforms, nNOS and eNOS, are controlled by calcium influx, which generates a burst of 10–50 nM NO lasting only minutes (61). However, phosphorylation of eNOS removes the calcium dependence and prolongs the production of NO (16, 45). The inducible form iNOS has always been assumed to generate only high concentrations of NO for prolonged periods. As discussed earlier, the local microenvironment of activated murine macrophages can be mimicked using millimolar concentrations of NO donors. In addition, NO flux in iNOS-expressing cells can range from high (300 nM) to low (<100 nM) levels, depending on the cytokine combination used for induction (13). These observations suggest that *in vivo* NO fluxes are controlled specifically and correlate with the three

distinct thresholds previously described (13, 64). In addition to its rate of synthesis and NO concentration, the biologic outcome also depends on cellular consumption and the redox environment. These discrete NO-signaling profiles suggest that NO concentration can be finely tuned to elicit different biologic responses, which has recently been extended by studies identifying an NO biphasic angiogenic response.

REGULATION OF ANGIOGENESIS THROUGH NO/TSP1 CROSS-TALK

Angiogenesis has emerged as a major target for the treatment of cancer (15, 19). Whereas numerous reports have shown both positive and negative effects of NO on angiogenesis (10), other studies have identified eNOS as a central regulator of the angiogenic process (8, 18, 58). Toward this end, it was recently shown that the regulatory event critical to the promotion of angiogenesis involves the phosphorylation of eNOS, which alters the enzyme to a calcium-independent state, resulting in the continuous flux of NO (18). Interestingly, many effects of various growth factors that regulate the induction of angiogenesis, such as VEGF, angiopoietin-2, and estrogen are all mediated through the phosphorylation of eNOS at serine 1179 (7, 14, 16, 28, 63), which activates eNOS for the production of NO. In contrast, the antiangiogenic agents, endostatin and somatostatin, both activate PP2A, an enzyme that dephosphorylates eNOS at serine 1179 (21, 67). Taken together, these observations are supportive of a central role of eNOS and NO generation during angiogenic response.

The regulation of angiogenesis involves a tight balance between pro- and antiangiogenic factors. Although NO has been shown to mediate the response of proangiogenic molecules, little is known of its role in the response(s) of antiangiogenic factors. Thrombospondins 1 (TSP1) and TSP2 are potent antiangiogenic matricellular proteins that exert diverse effects on several angiogenic cell responses including cell proliferation, adhesion, migration, and survival. TSP1 and TSP2 have been found to contribute to immune response, and they also function as tumor suppressors (37, 39, 44, 55). All of these effects arise from interaction of the whole protein or specific domains with various cell-surface receptors. Recently a unique cross-talk relation between NO and TSP1 has been demonstrated, which modulates angiogenic responses of vascular cells to NO, as shown in Fig. 1 (30). The identification of this important reciprocal relation between NO and TSP1 stems from the observation of enhanced NO-mediated endothelial cell migration in tissue samples taken from TSP1 null mice when compared with wild-type controls by using an explant model (30). This model used muscle biopsies implanted in type 1 collagen gel and incubated in growth medium for 10 days. Vascular cell (endothelial and smooth muscle) migration through the extracellular matrix was quantified by microscopically measuring the farthest distance of cell migration from the biopsy sample border. This method provides a rapid and reproducible assessment of vascular cell migration and matrix invasion, as hallmarks of an angiogenic response, as well as a means to test multiple pharmacologic agents in either a wound-healing or tumor-driven model of angiogenesis.

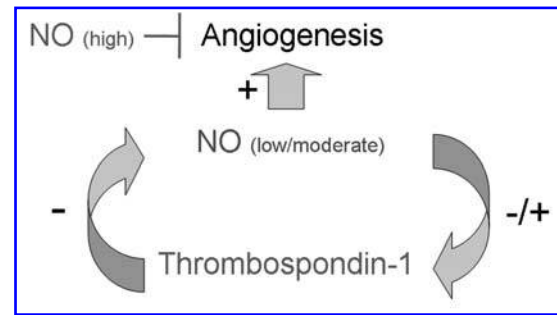


FIG. 1. Regulation of angiogenic response through NO/TSP1 crosstalk. Low levels of NO induce angiogenesis and downregulate TSP1. NO-induced angiogenic response is suppressed by increases in TSP1.

The explant model has shown that 100 μM DETA/NO ($t_{1/2}$ = 24 h) modestly enhanced endothelial cell migration in tissue from wild-type animals, whereas it dramatically accelerated vascular cell migration in tissue from TSP1-null animals. These results indicate that TSP1 is involved in the control of NO-stimulated angiogenic responses. In addition, vascular cell migration was found to respond to the NO donor DETA/NO in a biphasic manner; low doses (0.1–10 μM) enhanced cell migration, and high doses (>100–1,000 μM) attenuated the response. Similarly, supplemental L-arginine, a substrate of eNOS, increased vascular cell migration. However, in the presence of the NOS inhibitor L-NAME, the explant angiogenic response was completely abrogated. These results demonstrate that endogenous NOS activity and low doses of the NO donor stimulate an angiogenic response characterized by vascular cell invasion of and migration through extracellular collagen matrix. In contrast, higher concentrations of NO, consistent with that produced in the vicinity of activated macrophages, attenuated the angiogenic response (30).

TSP1 LIGATION OF CD36 INHIBITS THE NO ANGIOGENIC RESPONSE

An angiogenic response involves mobilization of an otherwise quiescent vascular network and changes in vascular cell adhesion, proliferation, and migration. *In vitro* analysis of each of these specific cellular responses demonstrated that both exogenous and endogenous TSP1 potentially inhibited the stimulatory effects of low-dose NO (30). More important, low-dose exogenous NO dramatically increased the sensitivity of endothelial cells to the inhibitory effects of exogenous TSP1. Although TSP1 at high doses ($\geq 10 \mu\text{g/ml}$) directly inhibits endothelial cell proliferation, adhesion, and migration, low-dose NO treatment ($\leq 10 \mu\text{M}$) increased the inhibitory potency of TSP1 by $\geq 1,000$ -fold. Further, these inhibitory effects were mimicked by a recombinant fragment of TSP1 containing its type 1 repeats. The major antiangiogenic effects of TSP1 have been mapped to this domain (66) and involve, in part, ligation of CD36 (9, 29). Interestingly, antibody ligation of CD36 also inhibited NO-driven endothelial cell responses in every angiogenic assay tested. These results

are particularly relevant because derivatives of TSP1 peptide sequences that act through the CD36 receptor have completed phase I clinical trials (23, 25) and are currently in phase II clinical trials for patients with renal cell carcinoma and soft tissue sarcomas (11).

On a molecular level, NO stimulates soluble guanylate cyclase, resulting in increased synthesis of intracellular cGMP. Significantly, Isenberg *et al.* (30) demonstrated that exogenous TSP1 prevented the NO-driven flux of intracellular cGMP, summarized in Fig. 2. CD36 antibody ligation and recombinant type 1 repeats of TSP1 also blocked NO-induced cGMP accumulation, implicating CD36 in this signaling response. Comparison of wild-type and TSP1-null mouse endothelial cell responses to NO showed that endogenous TSP1 also inhibits cGMP accumulation. This differential response to NO was not sensitive to inhibiting cGMP phosphodiesterases, indicating that TSP1 and CD36 signaling regulates synthesis rather than degradation of cGMP. This result supports the idea that sGC is a primary, although not the only, target for TSP1. It was also found that vascular cell responses, driven by 8-Br-cGMP, were inhibited by exogenous TSP1, indicating that at least one target for TSP1 exists downstream from sGC. These results suggest a new and important therapeutic paradigm for altering tumor-driven angiogenic responses based on dose dependence of NO flux and treatment with peptides or mimetics of TSP1 that act through CD36.

SIGNALING CASCADES AND NO PROLIFERATIVE RESPONSE IN ENDOTHELIAL CELLS

CD36-dependent signaling by TSP1 mediates inhibition of endothelial cell responses to NO and involves sGC as well as undefined targets downstream of cGMP. The identification of such targets may lead to new therapeutic strategies to control pathologic angiogenesis. Further understanding of the significance of this CD36 pathway in regulating NO-dependent angiogenic responses was achieved by examining the signaling profiles of key regulatory proteins in endothelial cells (51). In contrast to the response of MCF-7 cells, NO released by DETA/NO did not stabilize HIF-1 α in human umbilical vein endothelial cells (HUVECs), which could have been an effect of timing and donor specificity. However, the levels of p-

ser15-P53 increased in a dose-dependent manner with the highest induction (approximately sixfold above control) occurring at 1,000 μ M DETA/NO. Interestingly, an inverse relation between the levels of pERK and the MAPK phosphatase MKP-1 was found. Compared with control, increases in pERK followed a bell-shaped response, as low DETA/NO concentration (10 μ M) increased pERK levels whereas higher doses decreased phosphorylation. In contrast, MKP-1 levels decreased to a minimum in response to low-dose NO (10 μ M) and then rebounded to levels greater than twofold beyond control at high doses. These alterations in MAPK proteins and pP53 indicate that low-dose NO favors proliferation, whereas high doses are inhibitory, and in this respect are similar to the NO signaling profiles reported in MCF7 tumor cells (51, 64).

Endothelial cells, although known to synthesize and respond to TSP1, were found to respond to low-dose NO with a 50% decrease in expression of TSP1 (51), consistent with previous observations in kidney mesangial cells (68). As donor concentration increased, a modest reaccumulation of secreted TSP1 was observed (51). These findings suggest that NO-mediated proangiogenic responses involve the downregulation of TSP1. The presence of ODQ (guanylate cyclase inhibitor) or U0126 (MEK inhibitor) abated the NO-induced suppression of TSP1, as well as the proliferative response of HUVECs, indicating that the proangiogenic effects of NO are both cGMP and pERK dependent. Interestingly, exogenous TSP1 also inhibited NO-induced ERK phosphorylation, suggesting that in addition to cGMP dependence, the NO/TSP1 cross-talk is mediated, at least in part, through ERK phosphorylation; this is summarized in Fig. 3 (51).

RELEVANCE OF NO/TSP CROSSTALK

The role of NO in cancer depends on the temporal and spatial aspects of NO flux and changing cellular responses to NO during disease progression, which combine to yield either positive or negative outcomes for the tumor and host (74). Despite this complexity, a rationale for the duality of NO is beginning to emerge from the demonstration of the NO dose de-

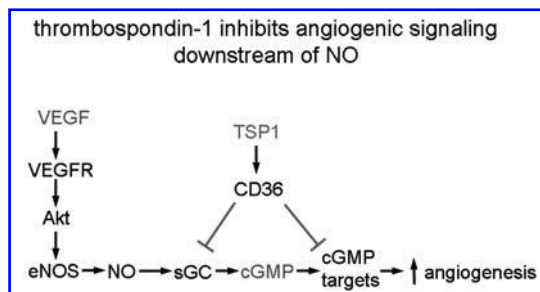


FIG. 2. Thrombospondin-1 suppression of NO-induced angiogenic response involves CD36 inhibition of sGC.

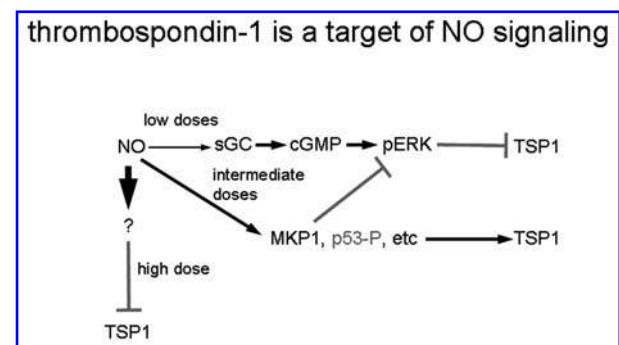


FIG. 3. Low fluxes of NO downregulate TSP1 in a sGC- and pERK-dependent manner, whereas moderate fluxes of NO are associated with P53 phosphorylation, MKP1 phosphatase induction, and modest reaccumulation of TSP1.

pendence of specific cellular and molecular targets in tumors (64). Further understanding has been derived from recent reports of the importance of redox flux in hypersensitizing vascular cells to the antiangiogenic effects of TSP1, a process involving cGMP and pERK (30, 51). An NO flux of as little as 2 nM was sufficient to induce vascular cell responses to picomolar amounts of exogenous TSP1, as manifested by inhibition of cellular adhesion, proliferation, and migration. A similar biphasic angiogenic response to the NO donor SNAP has been reported in microvascular endothelial cells, as low doses (0.1–0.3 mM) significantly enhanced cell migration, adhesion, and ERK phosphorylation, whereas higher doses (0.5–4 mM) attenuated these responses (33). Estradiol, a known mediator of tumor angiogenesis as well as eNOS activation, induced both proliferative and migration responses of HUVECs, which required ERK phosphorylation and TSP1 suppression (69, 59). These reports identify TSP1, cGMP, and ERK as regulators of NO-induced angiogenic response. Multiple approaches are currently being developed for the application of both TSP1 and TSP2 in cancer therapy (40, 70). Low-dose chemotherapy, also known as antiangiogenic chemotherapy or metronomic dosing, involves the optimization of the effects of cytotoxic drugs by administering them continuously at low, nontoxic doses (22, 40). Low-dose chemotherapy appears to provide a promising new approach because the targeted endothelial cells within the tumor bed are genetically stable and are therefore at a reduced risk of developing drug resistance, and low dosage produces significantly fewer side effects because of selectivity of endothelial cells (40). Toward this end, a recent report has shown that TSP1 secreted from the tumor microenvironment mediated the antiangiogenic and tumor growth-suppressive effects of low-dose cyclophosphamide (22). Evidence of endothelial cell selectivity was demonstrated by a reduction in CD31-positive vasculature (endothelial cell marker) in the tumors of cyclophosphamide-treated animals. Moreover, these antiangiogenic and tumor-suppressive responses did not occur with tumor cells that did not express TSP1 (22). These observations and the reports previously discussed, which demonstrated the involvement of NO generated by eNOS in tumor progression and metastasis (31, 34, 38), strengthen the potential relevance of the NO/TSP1 cross-talk relation (30, 51) in tumor biology. Taken together, these reports suggest that the elucidation of temporal and concentration-dependent redox pathways that drive tumor progression will facilitate the identification of novel therapeutic approaches for the treatment of cancer.

ACKNOWLEDGMENTS

This research was supported by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research.

ABBREVIATIONS

cGMP, cyclic guanosine monophosphate; DETA/NO, diethyltri-amine NONOate; HIF, hypoxia inducible factor; IL-

8, interleukin-8; MAPK, mitogen-activated protein kinase; NO, nitric oxide; pERK, phosphorylated extracellular regulated kinase; sGC, soluble guanylyl cyclase; NOS, nitric oxide synthase; RNOS, reactive nitrogen oxide species; Sper/NO, spermine NONOate; TNF- α , tumor necrosis factor- α ; TSP1, thrombospondin-1; VEGF, vascular endothelial growth factor.

REFERENCES

1. Ambs S, Bennett WP, Merriam WG, Ogunfusika MO, Oser SM, Harrington AM, Shields PG, Felley-Bosco E, Hussain SP, and Harris CC. Relationship between p53 mutations and inducible nitric oxide synthase expression in human colorectal cancer. *J Natl Cancer Inst* 91: 86–88, 1999.
2. Baker LH, Demetri GD, Mendelson DS, Rowinsky EK, McKeegan EM, Knight RA, Carlson DM, and Lobell M. A randomized phase 2 study of the thrombospondin-mimetic peptide ABT-510 in patients with advanced soft tissue sarcoma (STS). *J Clin Oncol* 23: 819S–819S.
3. Brune B. The intimate relation between nitric oxide and superoxide in apoptosis and cell survival. *Antiox Redox Signal* 7: 497–507, 2005.
4. Brune B and Mohr S. Protein thiol modification of glyceraldehyde-3-phosphate dehydrogenase and caspase-3 by nitric oxide. *Curr Protein Pept Sci* 2: 61–72, 2001.
5. Brune B, von Knethen K, and Sandau KB. Nitric oxide and its role in apoptosis. *Eur J Pharm* 351: 261–272, 1998.
6. Chen GG, Lau WY, Lai PBS, Chun YS, Chak ECW, Leung BCS, Lam IKY, Lee JFY, and Chui AKK. Activation of Kupffer cells inhibits tumor growth in a murine model system. *Int J Cancer* 99: 713–720, 2002.
7. Chen JX, Lawrence ML, Cunningham G, Christman BW, and Meyrick B. HSP90 and Akt modulate Ang-1-induced angiogenesis via NO in coronary artery endothelium. *J Appl Physiol* 96: 612–620, 2004.
8. Cooke JP. NO and angiogenesis. *Atherosclerosis Suppl* 4: 53–60, 2003.
9. Dawson DW, Pearce SFA, Zhong R, Silverstein RL, Frazier WA, and Bouck NP. CD36 mediates the in vitro inhibitory effects of thrombospondin1 on endothelial cells. *J Cell Biol* 138: 707–717, 1997.
10. Duda DG, Fukumura, D, and Jain RK. Role of eNOS in neovascularization: NO for endothelial progenitor cells. *Trends Mol Med* 10: 143–145, 2004.
11. Ebbinghaus SW, Hussain M, Tannir NM, Gordon MS, Desai AA, Knight RA, Carlson DM, Figlin RA. A randomized phase 2 study of the thrombospondin-mimetic peptide ABT-510 in patients with previously untreated advanced renal cell carcinoma. *J Clin Oncol* 23: 404S–404S, 2005.
12. Ekmekcioglu S, Tang CH, and Grimm EA. NO news is not necessarily good news in cancer. *Curr Cancer Drug Targets* 5: 103–115, 2005.
13. Espey MG, Miranda KM, Pluta, RM, and Wink DA. Nitrosative capacity of macrophages is dependent on nitric oxide synthase induction signals. *J Biol Chem* 275: 11341–11347, 2000.

14. Florian M, Lu Y, Angle M, and Magder S. Estrogen induced changes in Akt-dependent activation of endothelial nitric oxide synthase and vasodilation. *Steroids* 69: 637–645, 2004.
15. Folkman J. Fundamental concepts of the angiogenic process. *Curr Mol Med* 3: 643–651, 2003.
16. Fulton D, Fontana J, Sowa G, Gratton JP, Lin M, Li KX, Michell B, Kemp B, Rodman D, and Sessa WC. Localization of endothelial nitric oxide synthase phosphorylated on serine 1179 and nitric oxide in Golgi and plasma membrane defines the existence of two pools of active enzyme. *J Biol Chem* 277: 4277–4284, 2002.
17. Fulton D, Gratton JP, McCabe TJ, Fontana J, Fujio Y, Walsh K, Franke TF, Papapetropoulos A, and Sessa WC. Regulation of endothelium-derived nitric oxide production by the protein kinase Akt. *Nature* 399: 597–601, 1999.
18. Fulton D, Gratton JP, and Sessa WC. Post-translational control of endothelial nitric oxide synthase: Why isn't calcium/calmodulin enough? *J Pharmacol Exp Ther* 299: 818–824, 2001.
19. Gasparini G, Longo R, Fanelli M, and Teicher BA. Combination of antiangiogenic therapy with other anticancer therapies: Results, challenges, and open questions. *J Clin Oncol* 23: 1295–311, 2005.
20. Goodman J, Hofseth LJ, Hussain SP, and Harris CC. Nitric oxide and p53 in cancer-prone chronic inflammation and oxyradical overload disease. *Environ Mol Mutagen* 44: 3–9, 2004.
21. Greif DM, Kou R, and Michel T. Site-specific dephosphorylation of endothelial nitric oxide synthase by protein phosphatase 2A: Evidence for crosstalk between phosphorylation sites. *Biochemistry* 41: 15845–15853, 2002.
22. Hamano Y, Sugimoto H, Soubasakos MA, Kieran M, Olsen BR, Lawler J, Sudhakar A, and Kalluri R. Thrombospondin1 associated with tumor microenvironment contributes to low-dose cyclophosphamide-mediated endothelial cell apoptosis and tumor growth suppression. *Cancer Res* 64: 1570–1574, 2004.
23. Haviv F, Bradley MF, Kalvin DM, Schneider AJ, Davidson DJ, Majest SM, McKay LM, Haskell CJ, Bell RL, Nguyen B, Marsh KC, Surber BW, Uchic JT, Ferrero J, Wang YC, Leal J, Record RD, Hodde J, Badylak SF, Lesniewski RR, and Henkin J. Thrombospondin1 mimetic peptide inhibitors of angiogenesis and tumor growth: Design, synthesis, and optimization of pharmacokinetics and biological activities. *J Med Chem* 48: 2838–2846, 2005.
24. Hibbs JB, Taintor RR, and Vavrin Z. Macrophage cytotoxicity: Role for L-arginine deiminase and imino nitrogen oxidation to nitrite. *Science* 235: 473–476, 1987.
25. Hoekstra R, de Vos FYFL, Eskens FALM, Gietema JA, van der Gaast A, Groen HJM, Knight RA, Carr RA, Humerickhouse RA, Verweij J, and de Vries EGE. Phase I safety, pharmacokinetic, and pharmacodynamic study of the thrombospondin1-mimetic angiogenesis inhibitor ABT-510 in patients with advanced cancer. *J Clin Oncol* 23: 5188–5197, 2005.
26. Hofseth LJ, Saito S, Hussain SP, Espey MG, Miranda KM, Araki Y, Jhappan C, Higashimoto Y, He P, Linke SP, Quezado MM, Zurer I, Rotter V, Wink DA, Appella E, and Harris CC. Nitric oxide induced cellular stress and P53 activation in chronic inflammation. *Proc Natl Acad Sci USA* 100: 143–148, 2003.
27. Hussain SP, Trivers GE, Hofseth LJ, He P, Shaikh I, Mechanic LE, Doja S, Jiang W, Subleski J, Shorts L, Haines D, Laubach VE, Wiltout RH, Djurickovic D, and Harris CC. Nitric oxide, a mediator of inflammation, suppresses tumorigenesis. *Cancer Res* 64: 6849–6853, 2004.
28. Igarashi J and Michel T. Sphingosine1-phosphate and isoform-specific activation of phosphoinositide 3-kinase. *J Biol Chem* 276: 36281–36288, 2001.
29. Iruela-Arispe LM, Lombardo M, Krutzsch HC, Lawler J, and Roberts DD. Inhibition of angiogenesis by thrombospondin1 is mediated by two independent regions within the type 1 repeats. *Circulation* 100: 1423–1431, 1999.
30. Isenberg JS, Ridnour LA, Perruccio EM, Espey MG, Wink DA, and Roberts DD. Thrombospondin1 inhibits endothelial cell responses to nitric oxide in a cGMP-dependent manner. *Proc Natl Acad Sci USA* 102: 13141–13146, 2005.
31. Jadeski LC, Chakraborty C, and Lala PK. Nitric oxide-mediated promotion of mammary tumour cell migration requires sequential activation of nitric oxide synthase, guanylate cyclase and mitogen-activated protein kinase. *Int J Cancer* 106: 496–504, 2003.
32. Jenkins DC, Charles IG, Thomsen LL, Moss DW, Holmes LS, Baylis SA, Rhodes P, Westmore K, Emson PC, and Moncada S. Roles of nitric oxide in tumor growth. *Proc Natl Acad Sci USA* 92: 4392–4396, 1995.
33. Jones MK, Tsugawa K, Tarnawski AS, and Baatar D. Dual actions of nitric oxide on angiogenesis: Possible roles of PKC, ERK, and AP-1. *Biochem Biophys Res Commun* 318: 520–528, 2004.
34. Kashiwagi S, Izumi Y, Gohongi T, Demou ZN, Xu L, Huang PL, Buerk DG, Munn LL, Jain RK, and Fukumura D. NO mediates mural cell recruitment and vessel morphogenesis in murine melanomas and tissue-engineered blood vessels. *J Clin Invest* 115: 1816–1827, 2005.
35. Kim SJ, Hwang SG, Shin DY, Kang SS, and Chun JS. p38 kinase regulates nitric oxide induced apoptosis of articular chondrocytes by accumulating p53 via NFkB-dependent transcription and stabilization by serine 15 phosphorylation. *J Biol Chem* 277: 33501–33508, 2002.
36. Kim YM, Talanian RV, and Billiar TR. Nitric oxide inhibits apoptosis by preventing increases in caspase-3-like activity via two distinct mechanisms. *J Biol Chem* 272: 31138–31148, 1997.
37. Kuznetsova SA and Roberts DD. Functional regulation of T lymphocytes by modulatory extracellular matrix proteins. *Int J Biochem Cell Biol* 36: 1126–1134, 2004.
38. Lala PK and Orlucevic A. Role of nitric oxide in tumor progression: lessons from experimental tumors. *Cancer Metastasis Rev* 17: 91–106, 1998.
39. Lawler J. Thrombospondin1 as an endogenous inhibitor of angiogenesis and tumor growth. *J Cell Mol Med* 6: 1–12, 2002.
40. Lawler J and Detmar M. Tumor progression: the effects of thrombospondin-1 and -2. *Int J Biochem Cell Biol* 36: 1038–1045, 2004.
41. Le X, Wei D, Huang S, Lancaster JR Jr, and Xie K. Nitric oxide synthase II suppresses the growth and metastasis of

- human cancer regardless of its up-regulation of pro-tumor factors. *Proc Natl Acad Sci U S A* 102: 8758–8763, 2005.
42. Lechner M, Lirk P, and Rieder J. Inducible nitric oxide (iNOS) in tumor biology: The two sides of the same coin. *Semin Cancer Biol* 15: 277–289, 2005.
 43. Li J, Billiar TR, Talanian RV, and Kim YM. Nitric oxide reversibly inhibits seven members of the caspase family via S-nitrosylation. *Biochem Biophys Res Commun* 240: 419–424, 1997.
 44. Li Z, He L, Wilson KE, and Roberts DD. Thrombospondin1 inhibits TCR-mediated T lymphocyte early activation. *J Immunol* 166: 2427–2436, 2001.
 45. McCabe TJ, Fulton D, Roman LJ, and Sessa WC. Enhanced electron flux and reduced calmodulin dissociation may explain “calcium-independent” eNOS activation by phosphorylation. *J Biol Chem* 275: 6123–6128, 2000.
 46. Meßmer UK, Ankarcrona M, Nicotera P, and Brüne B. P53 Expression in nitric oxide-induced apoptosis. *FEBS Lett* 355: 23–26, 1994.
 47. Metzen E, Zhou J, Jelkmann W, Fandrey J, and Brüne B. Nitric oxide impairs normoxic degradation of HIF-1 α by inhibition of prolyl hydroxylases. *Mol Biol Cell* 14: 3470–3481, 2003.
 48. Mohr S, Zech B, Lapetina EG, and Brüne B. Inhibition of caspase-3 by S-nitrosation and oxidation caused by nitric oxide. *Biochem Biophys Res Commun* 238: 387–391, 1997.
 49. Palmer L, Gaston B, and Johns RA. Normoxic stabilization of hypoxia-inducible factor-1 expression and activity: Redox-dependent effect of nitrogen oxides *Mol Pharm* 58: 1197–1203, 2000.
 50. Pervin S, Singh R, Freije WA, and Chaudhuri G. MKP1-induced dephosphorylation of extracellular signal-regulated kinase is essential for triggering nitric oxide-induced apoptosis in human breast cancer cell lines: Implications in breast cancer. *Cancer Res* 63: 8853–8860, 2003.
 51. Ridnour LA, Isenberg JS, Espey MD, Thomas DD, Roberts DD, and Wink DA. Nitric oxide regulates angiogenesis through a functional switch involving thrombospondin1. *Proc Natl Acad Sci U S A* 102: 13147–13152, 2005.
 52. Ridnour LA, Oberley TD, and Oberley LW. Tumor suppressive effects of MnSOD overexpression may involve imbalance in peroxide generation versus peroxide removal. *Antiox Redox Signal* 6: 501–512, 2004.
 53. Ridnour LA, Sim JE, Choi J, Dickinson DA, Forman HJ, Ahmad IM, Coleman MC, Hunt CR, Goswami PC, and Spitz DR. Nitric oxide-induced resistance to hydrogen peroxide stress is a glutamate cysteine ligase activity-dependent process. *Free Radic Biol Med* 38: 1361–1371, 2005.
 54. Ridnour LA, Thomas DD, Mancardi D, Espey MG, Miranda KM, Paolocci N, Feelisch M, Fukuto J, and Wink DA. The chemistry of nitrosative stress induced by nitric oxide and reactive nitrogen oxide species: Putting perspective on stressful biological situations. *Biol Chem* 385: 1–10, 2004.
 55. Roberts DD. Regulation of tumor growth and metastasis by thrombospondin1. *FASEB J* 10: 1183–1191, 1996.
 56. Schmid T, Zhou J, and Brüne B. HIF-1 and P53: Communication of transcription factors under hypoxia. *J Cell Mol Med* 4: 423–431, 2004.
 57. Schneiderhan N, Budde A, Zhang Y, and Brüne B. Nitric oxide induces phosphorylation of P53 and impairs nuclear export. *Oncogene* 22: 2857–2868, 2003.
 58. Schwentker A, Vodovotz Y, Weller R, and Billiar TR. Nitric oxide and wound repair: Role of cytokines? *Nitric Oxide* 7: 1–10, 2002.
 59. Sengupta K, Banerjee S, Saxena NK, and Banerjee SK. Thrombospondin1 disrupts estrogen-induced endothelial cell proliferation and migration and its expression is suppressed by estradiol. *Mol Cancer Res* 2: 150–158, 2004.
 60. Stuehr DJ. Structure-function aspects in the nitric oxide synthases. *Annu Rev Pharmacol Toxicol* 37: 339–359, 1997.
 61. Stuehr DJ, Santolini J, Wang ZQ, Wei CC, and Adak S. Update on mechanism and catalytic regulation in the NO synthases. *J Biol Chem* 279: 36167–36170, 2004.
 62. Sumbayev VV, Sandau KB, and Brüne B. Mesangial cells but not hepatocytes are protected against NO/O₂⁻ cogeneration: Mechanistic considerations. *Eur J Pharm* 444: 1–11, 2002.
 63. Takahashi S and Mendelsohn ME. Synergistic activation of endothelial nitric oxide synthase (eNOS) by HSP90 and Akt. *J Biol Chem* 278: 30821–30827, 2003.
 64. Thomas DD, Espey MG, Ridnour LA, Hofseth LJ, Mancardi D, Harris CC, and Wink DA. Hypoxic inducible factor 1 α , extracellular signal-regulated kinase, and P53 are regulated by distinct threshold concentrations of nitric oxide. *Proc Natl Acad Sci U S A* 101: 8894–8899, 2004.
 65. Thomsen LL and Miles DW. Role of nitric oxide in tumour progression: lessons from human tumours. *Cancer Metastasis Rev* 17: 107–118, 1998.
 66. Tolsma SS, Volpert OV, Good DJ, Frazier WA, Polverini PJ, and Bouck N. Peptides derived from two separate domains of the matrix protein thrombospondin1 have anti-angiogenic activity. *J Cell Biol* 122: 497–511, 1993.
 67. Urbich C, Reissner A, Chavakis E, Dernbach E, Haendeler J, Fleming I, Zeiher AM, Kaszkin M, and Dimmeler S. Dephosphorylation of endothelial nitric oxide synthase contributes to the anti-angiogenic effects of endostatin. *FASEB J* 16: 706–708, 2002.
 68. Wang S, Shiva S, Poczatek MH, Darley-Usmar V, and Murphy-Ullrich JE. Nitric oxide and cGMP-dependent protein kinase regulation of glucose-mediated thrombospondin 1-dependent transforming growth factor- β activation in mesangial cells. *J Biol Chem* 277: 9880–9888, 2002.
 69. Weiner CP, Lizasoain I, Baylis SA, Knowles RG, Charles IG, and Moncada S. Induction of calcium-dependent nitric oxide synthases by sex hormones. *Proc Natl Acad Sci U S A* 91: 5212–5216, 1994.
 70. Westphal HR. Technology evaluation: ABT-510, Abbott. *Curr Opin Mol Ther* 6: 451–459, 2004.
 71. Wink DA, Cook JA, Pacelli R, Liebmann J, Krishna MC, and Mitchell JB. Nitric oxide (NO) protects against cellular damage by reactive oxygen species. *Toxicol Lett* 82: 83:221–226, 1995.
 72. Wink DA and Mitchell JB. Chemical biology of nitric oxide: insights into regulatory, cytotoxic, and cytoprotective mechanisms of nitric oxide. *Free Radic Biol Med* 25: 434–456, 1998.

73. Wink DA and Mitchell JB. Nitric oxide and cancer: an introduction. *Free Radic Biol Med* 34: 951–954, 2003.
74. Wink DA, Vodovotz Y, Laval J, Laval F, Dewhirst MW, and Mitchell JB. The multifaceted roles of nitric oxide in cancer. *Carcinogenesis* 19: 711–721, 1998.
75. Xie K, Huang S, Dong Z, Gutman M, and Fidler IJ. Direct correlation between expression of endogenous inducible nitric oxide synthase and regression of M5076 reticulum cell sarcoma hepatic metastases in mice treated with liposomes containing lipopeptide CGP 31362. *Cancer Res* 55: 3123–3131, 1995.
76. Xie K, Huang S, Dong Z, Juang SH, Gutman M, Xie QW, Nathan C, and Fidler IJ. Transfection with the inducible nitric oxide synthase gene suppresses tumorigenicity and abrogates metastasis by K-1735 murine melanoma cells. *J Exp Med* 181: 1333–1343, 1995.
77. Xie K, Huang S, Dong Z, Juang SH, Wang Y, and Fidler IJ. Destruction of bystander cells by tumor cells transfected with inducible nitric oxide (NO) synthase gene. *J Natl Cancer Inst* 89: 421–427, 1997.
78. Zech B, Kohl R, von Knethen A, and Brune B. Nitric oxide donors inhibit formation of the Apaf-1/caspase-9 apoptosome and activation of caspases. *Biochem J* 371: 1055–1064, 2003.
79. Zech B, Wilm M, van Eldik R, and Brune B. Mass spectrometric analysis of nitric oxide-modified caspase-3. *J Biol Chem* 274: 20931–20936, 1999.
80. Zhou J and Brune B. NO and transcriptional regulation: from signaling to death. *Toxicology* 208: 223–233, 2005.
81. Zhou J, Schmid T, and Brune B. HIF-1 and P53 as targets of NO in affecting cell proliferation, death and adaptation. *Curr Mol Med* 4: 741–751, 2004.
82. Zurer I, Hofseth LJ, Cohen Y, Xu-Welliver M, Hussain SP, Harris CC, and Rotter V. The role of P53 in base excision repair following genotoxic stress. *Carcinogenesis* 25: 11–9, 2004.

Address reprint requests to:

Lisa A. Ridnour

Radiation Biology Branch

National Cancer Institute/National Institutes of Health

10 Center Drive

Building 10, Room B3-B35

Bethesda, MD 20892

E-mail: ridnourl@mail.nih.gov

Date of first submission to ARS Central, January 24, 2006;
date of acceptance, February 7, 2006.

This article has been cited by:

1. Wagner L. Batista , Fernando T. Ogata , Marli F. Curcio , Rodrigo B. Miguel , Roberto J. Arai , Alisson L. Matsuo , Miriam S. Moraes , Arnold Stern , Hugo P. Monteiro . S-Nitrosoglutathione and Endothelial Nitric Oxide Synthase-Derived Nitric Oxide Regulate Compartmentalized Ras S-Nitrosylation and Stimulate Cell Proliferation. *Antioxidants & Redox Signaling*, ahead of print. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)] [[Supplemental material](#)]
2. Shan Yu, Lin Jia, Yan Zhang, Dinglan Wu, Zhenyu Xu, Chi-Fai Ng, Kenneth K.W. To, Yu Huang, Franky L. Chan. 2012. Increased expression of activated endothelial nitric oxide synthase contributes to antiandrogen resistance in prostate cancer cells by suppressing androgen receptor transactivation. *Cancer Letters* . [[CrossRef](#)]
3. Päivi Rahkola-Soisalo, Hanna Savolainen-Peltonen, Mervi Väisänen-Tommiska, Ralf Butzow, Olavi Ylikorkala, Tomi S. Mikkola. 2012. High-risk human papillomavirus-induced expression of endothelial and inducible nitric oxide synthase in human uterine cervix. *Annals of Medicine* 1-6. [[CrossRef](#)]
4. Jack Henkin, Olga V Volpert. 2011. Therapies using anti-angiogenic peptide mimetics of thrombospondin-1. *Expert Opinion on Therapeutic Targets* 1-18. [[CrossRef](#)]
5. Kewei Wang, John J. Brems, Richard L. Gamelli, Ai-Xuan Holterman. 2011. iNOS/NO signaling regulates apoptosis induced by glycochenodeoxycholate in hepatocytes. *Cellular Signalling* **23**:10, 1677-1685. [[CrossRef](#)]
6. Simendra Singh, Alok K. Gupta. 2011. Nitric oxide: role in tumour biology and iNOS/NO-based anticancer therapies. *Cancer Chemotherapy and Pharmacology* **67**:6, 1211-1224. [[CrossRef](#)]
7. Qing-Ping Zeng, Ping-Zu Zhang. 2011. Artesunate mitigates proliferation of tumor cells by alkylating heme-harboring nitric oxide synthase. *Nitric Oxide* **24**:2, 110-112. [[CrossRef](#)]
8. A. Weyerbrock, S. Walbridge, J. E. Saavedra, L. K. Keefer, E. H. Oldfield. 2011. Differential effects of nitric oxide on blood-brain barrier integrity and cerebral blood flow in intracerebral C6 gliomas. *Neuro-Oncology* **13**:2, 203-211. [[CrossRef](#)]
9. Rouba Ali-Fehmi, Assaad Semaan, Sima Sethi, Haitham Arabi, Sudeshna Bandyopadhyay, Yaser R. Hussein, Michael P. Diamond, Ghasan Saed, Robert T. Morris, Adnan R. Munkarah. 2011. Molecular typing of epithelial ovarian carcinomas using inflammatory markers. *Cancer* **117**:2, 301-309. [[CrossRef](#)]
10. Evangelos Kogias, Nadja Osterberg, Brunhilde Baumer, Nikolaos Psarras, Christoph Koentges, Anna Papazoglou, Joseph E. Saavedra, Larry K. Keefer, Astrid Weyerbrock. 2011. Growth-inhibitory and chemosensitizing effects of the glutathione-S-transferase- γ -activated nitric oxide donor PABA/NO in malignant gliomas. *International Journal of Cancer* n/a-n/a. [[CrossRef](#)]
11. Keri Seymour, Xuan Han, Benjamin Sadowitz, Kristopher G. Maier, Vivian Gahtan. 2010. Differential effect of nitric oxide on thrombospondin-1-, PDGF- and fibronectin-induced migration of vascular smooth muscle cells. *The American Journal of Surgery* **200**:5, 615-619. [[CrossRef](#)]
12. J. M. Weiss, L. A. Ridnour, T. Back, S. P. Hussain, P. He, A. E. Maciag, L. K. Keefer, W. J. Murphy, C. C. Harris, D. A. Wink, R. H. Wiltout. 2010. Macrophage-dependent nitric oxide expression regulates tumor cell detachment and metastasis after IL-2/anti-CD40 immunotherapy. *Journal of Experimental Medicine* **207**:11, 2455-2467. [[CrossRef](#)]
13. Monica G. Ferrini, Steve Rivera, Joanne Moon, Dolores Vernet, Jacob Rajfer, Nestor F. Gonzalez-Cadavid. 2010. The Genetic Inactivation of Inducible Nitric Oxide Synthase (iNOS) Intensifies Fibrosis and Oxidative Stress in the Penile Corpora Cavernosa in Type 1 Diabetes. *The Journal of Sexual Medicine* **7**:9, 3033-3044. [[CrossRef](#)]
14. Vinod Prabhu, C. Guruvayoorappan. 2010. Nitric oxide: pros and cons in tumor progression. *Immunopharmacology and Immunotoxicology* **32**:3, 387-392. [[CrossRef](#)]
15. Benjamin J. Vesper, Kim M. Elseth, Gabor Tarjan, G. Kenneth Haines, James A. Radosevich. 2010. Long-term adaptation of breast tumor cell lines to high concentrations of nitric oxide. *Tumor Biology* **31**:4, 267-275. [[CrossRef](#)]
16. Yu-Der Wen, Yun-Lung Ho, Rong-Jen Shiau, Jung-Kai Yeh, Jheng-Yu Wu, Wei-Lung Wang, Show-Jen Chiou. 2010. Synergistic antitumor effect of curcumin and dinitrosyl iron complexes for against melanoma cells. *Journal of Organometallic Chemistry* **695**:3, 352-359. [[CrossRef](#)]
17. Nicholas A. Pullen, Helen L. Fillmore. 2010. Induction of matrix metalloproteinase-1 and glioma cell motility by nitric oxide. *Journal of Neuro-Oncology* **96**:2, 201-209. [[CrossRef](#)]
18. Robert Cheng, Sharon Glynn, Wilmarie Flores-Santana, Christopher Switzer, Lisa Ridnour, David A. WinkNitric Oxide and Redox Inflammation in Cancer **4**, 157-182. [[CrossRef](#)]
19. Hao Hong, Jiangtao Sun, Weibo Cai. 2009. Multimodality imaging of nitric oxide and nitric oxide synthases. *Free Radical Biology and Medicine* **47**:6, 684-698. [[CrossRef](#)]

20. Paivi Rahkola, Tomi S. Mikkola, Olavi Ylikorkala, Mervi Vaisanen-Tommiska. 2009. Association between high risk papillomavirus DNA and nitric oxide release in the human uterine cervix. *Gynecologic Oncology* **114**:2, 323-326. [[CrossRef](#)]
21. Verena Fetz, Carolin Bier, Negusse Habtemichael, Robert Schuon, Andrea Schweitzer, Martin Kunkel, Knut Engels, Adorján F. Kovács, Sandra Schneider, Wolf Mann, Roland H. Stauber, Shirley K. Knauer. 2009. Inducible NO synthase confers chemoresistance in head and neck cancer by modulating survivin. *International Journal of Cancer* **124**:9, 2033-2041. [[CrossRef](#)]
22. Khosrow Kashfi. Anti-Inflammatory Agents as Cancer Therapeutics **57**, 31-89. [[CrossRef](#)]
23. Jolie Kiemlian Kwee, Diogo Gomes Luque, Ana Carolina dos Santos Ferreira, Flavia da Cunha Vasconcelos, Karina Lani Silva, Claudete Esteves Klumb, Raquel Ciuvalschi Maia. 2008. Modulation of reactive oxygen species by antioxidants in chronic myeloid leukemia cells enhances imatinib sensitivity through survivin downregulation. *Anti-Cancer Drugs* **19**:10, 975-981. [[CrossRef](#)]
24. Fabienne Peyrot, Claire Ducrocq. 2008. Potential role of tryptophan derivatives in stress responses characterized by the generation of reactive oxygen and nitrogen species. *Journal of Pineal Research* **45**:3, 235-246. [[CrossRef](#)]
25. V. Badireenath Konkimalla, Martina Blunder, Bernhard Korn, Shahid A. Soomro, Herwig Jansen, Wonsuk Chang, Gary H. Posner, Rudolf Bauer, Thomas Efferth. 2008. Effect of artemisinins and other endoperoxides on nitric oxide-related signaling pathway in RAW 264.7 mouse macrophage cells. *Nitric Oxide* **19**:2, 184-191. [[CrossRef](#)]
26. Anand Krishnan V. Iyer, Neelam Azad, Liying Wang, Yon Rojanasakul. 2008. Role of S-nitrosylation in apoptosis resistance and carcinogenesis. *Nitric Oxide* **19**:2, 146-151. [[CrossRef](#)]
27. Lisa A. Ridnour, Douglas D. Thomas, Christopher Switzer, Wilmarie Flores-Santana, Jeffrey S. Isenberg, Stefan Ambs, David D. Roberts, David A. Wink. 2008. Molecular mechanisms for discrete nitric oxide levels in cancer. *Nitric Oxide* **19**:2, 73-76. [[CrossRef](#)]
28. Mario E. Goetz, Andreas Luch. 2008. Reactive species: A cell damaging route assisting to chemical carcinogens. *Cancer Letters* **266**:1, 73-83. [[CrossRef](#)]
29. J. Bordini, D.O. Novaes, I.E. Borissevitch, B.T. Owens, P.C. Ford, E. Tfouni. 2008. Acidity and photolability of ruthenium salen nitrosyl and aquo complexes in aqueous solutions. *Inorganica Chimica Acta* **361**:8, 2252-2258. [[CrossRef](#)]
30. Elizabeth Anne Hillard, Fabiane Caxico de Abreu, Danielle Cristhina Melo Ferreira, Gérard Jaouen, Marília Oliveira Fonseca Goulart, Christian Amatore. 2008. Electrochemical parameters and techniques in drug development, with an emphasis on quinones and related compounds. *Chemical Communications* :23, 2612. [[CrossRef](#)]
31. Rosekeila Simões Nomelini, Lívia Carolina de Abreu Ribeiro, Beatriz Martins Tavares-Murta, Sheila Jorge Adad, Eddie Fernando Candido Murta. 2008. Production of Nitric Oxide and Expression of Inducible Nitric Oxide Synthase in Ovarian Cystic Tumors. *Mediators of Inflammation* **2008**, 1-7. [[CrossRef](#)]
32. Brandon G. Bentz, Neal D. Hammer, Brett Milash, Slobodanka Klein, David M. Burnett, James A. Radosevich, G. Kenneth Haines, III. 2007. The Kinetics and Redox State of Nitric Oxide Determine the Biological Consequences in Lung Adenocarcinoma. *Tumor Biology* **28**:6, 301-311. [[CrossRef](#)]